

Pirkle, 1977). One of these, batroxobin (from *Rothmans* venoms) has been cloned and sequenced (Itoh et al., 1987; 1988). Peptide paranitroaniline (pNA) Chemical: Biochemical Corp. and Sigma

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was made for the heparin-like sites. The reaction mixture was dialyzed into 50 mM Tris-HCl and for the pH 8.0 and applied to a 1.5 cm x 75 cm using ProPheArgNA and PheValArg. Sephadex G-75 gel filtration column. Flow: thrombin-like protease using ions pNA. An appropriate amount of substrate rate was 4.8 ml/hr with four fractions. Fractions 50-60 containing was added to give a total volume of 675 μ l collected/hr.

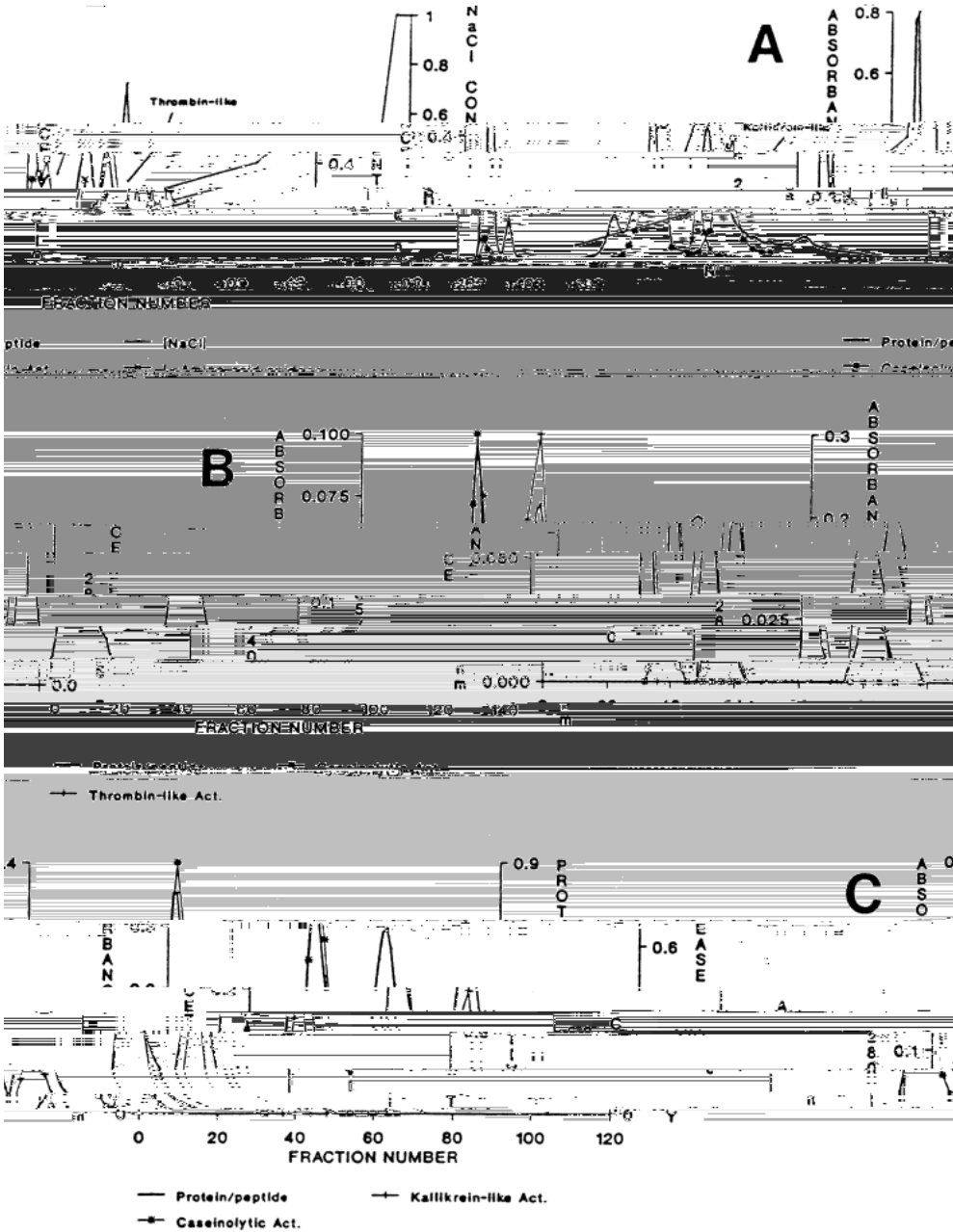


Fig. 1. Separation of thrombin-like protease from venous thrombolytic material. A. Thrombin-like protease was separated from venous thrombolytic material by ion exchange column. B. Elution profile of combined fractions from the thrombin-like protease peak chromatographed on a 1.5 cm x 75 cm Sephadex G-75 gel filtration column. C. Elution profile of combined fractions from the thrombin-like protease peak chromatographed on a 1.5 cm x 75 cm Sephadex G-75 gel filtration column. Thrombin-like activity was completely separated from caseinolytic activity.

Hydrolyzed
Peptide

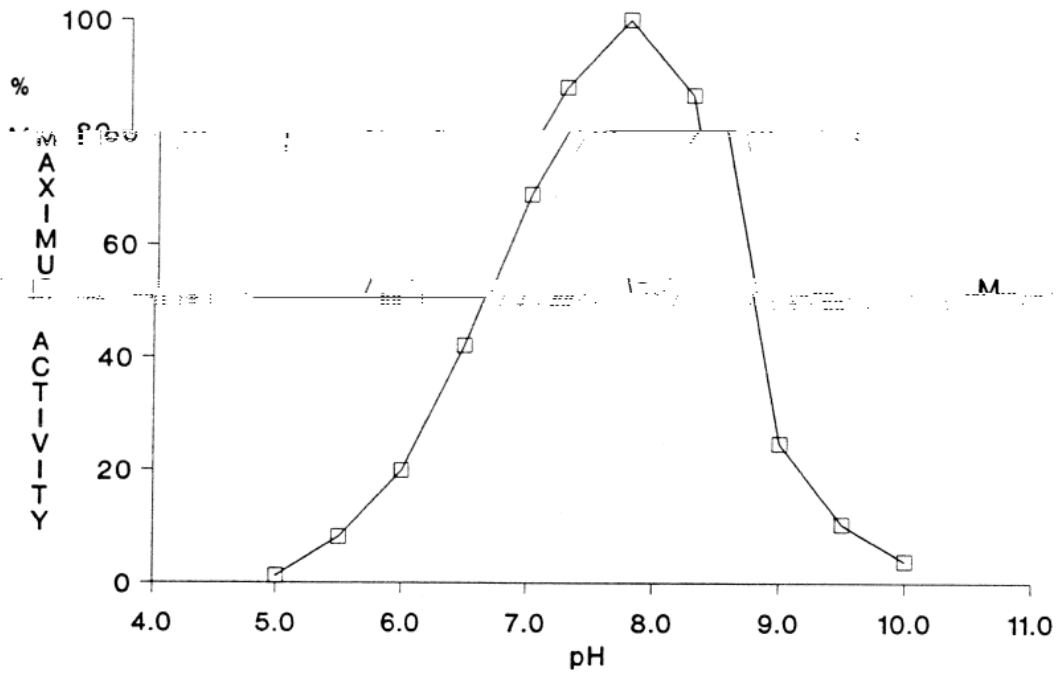


Figure 3. pH-profile for the hydrolysis of NBD-Casein. The activity is expressed as a percentage of the activity observed on highest activity observed (at pH 8.0).

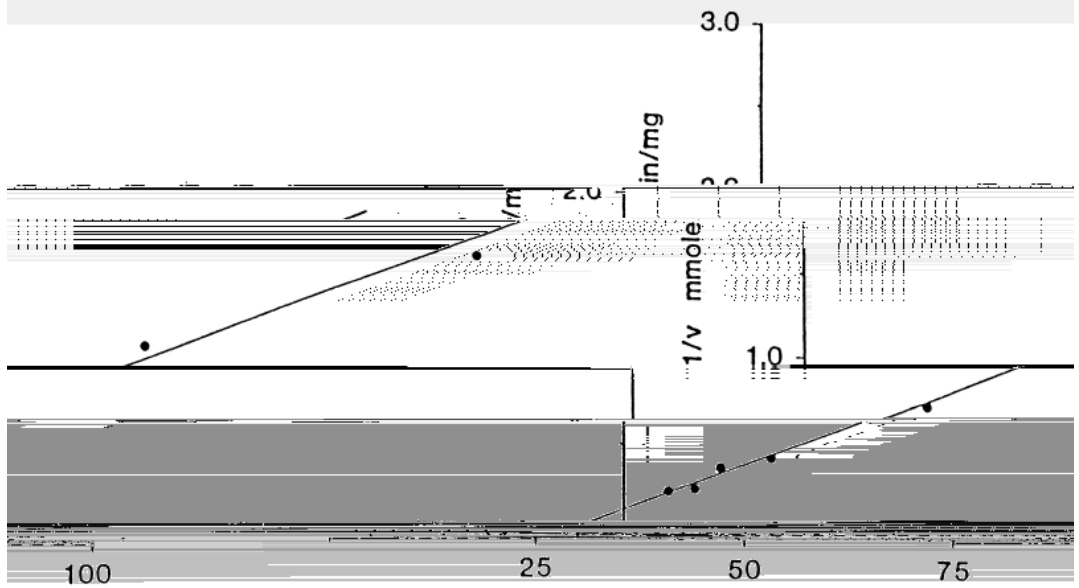


Figure 4. Double-reciprocal plot of $1/v$ vs. substrate concentration for the hydrolysis of NBD-Casein. $K_m = 22 \mu M$, $V_m = 1.413 \text{ } \mu\text{mol/min/mg}$.

The kallikrein III
had a K_m of 0.00125 mM and a V_{max} of 0.000125 mM. It showed a high degree of specificity, as
hydrolyzed to a significant extent had \equiv kallikrein substrate ProPheArg-pNA with

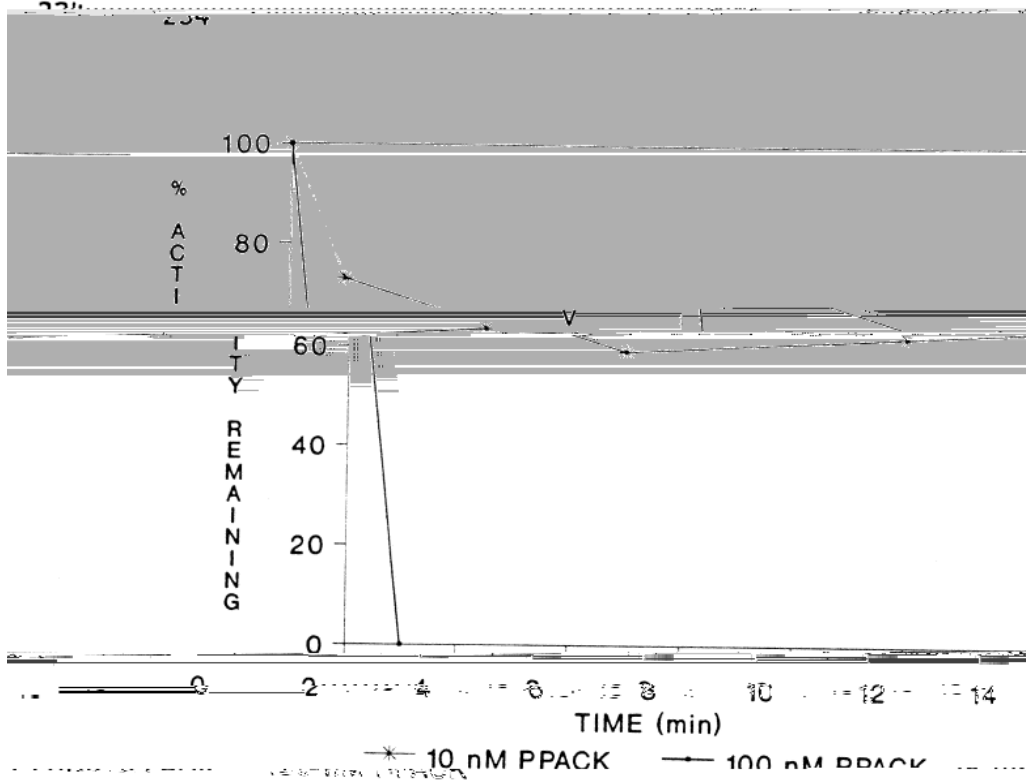


Figure 9
 Inhibition of vesicle-facilitated activity by PPACK. 100 nM PPACK resulted in the complete inhibition of ~0.5 nM enzyme within one minute.

It must be recalled to determine the kallikrein-like enzymes isolated from C. venen (Schwarz and Bröck, particular activities attributable to them (Schwarz and Bröck, 1965). The approach of venom isolated from *Bilis gabonica* venom (Vilicic, potential synergistic effects constituents. In order to understand the (et al., 1979).

biological role of venoms as biological

resemble the summed effects of isolated Components

rather than being Rattlesnake venoms have two venom components. Rather than being independent components, major biological functions related to treated as individual inde

they analyzed for their potentially interdependent effects. Zeller (1977) has made the (Klauber, 1956; Russell, 1980; Kard

achieved by biochemistry with the view that venoms primary importance and is acco

in these venoms (Lee, 1979; Tu, 1983). In

cause circulatory shock. In humans, locally impeding defense against snake bites is reportedly harmful (Russell, 1980). In prey animals, with a much (Ownby, 1982); and the synergistic action of this tissue reaction is likely accentuated by venom may be responsible for much interest to note that the enzymes interfering with hemostasis. It is of major importance to note that the enzymes interfering with hemostasis are major metalloproteinases. In juvenile snake venoms (Mackessy, 1993), and bites by smaller snakes rarely result in

Russell. Venom proteases may act as toxins local tissue necrosis (Russell, 1980).

the course of envenomation. In the blood, the thrombin-like enzymes have relatively specific activities of fibrinogen to both adult and juvenile venoms, and mainly the hydrolytic enzymes may also act as toxins may produce several different effects

and a precursor to the hormone cell cultures (Bar-Shavit et al. 1986 activities of

thrombin-like proteases may also have enzymes release active in the blood. The immediate action of these enzymes is via the release of potent fibrinogen. The relative abundance of as toxins is via the release of active and active two substances which appear to metabolites such as precursors and change unambiguously suggesting that a carbonic acid anhydrase enzyme is probably involved in the release of these metabolites.

venom neurotoxins). The toxic action of these metabolites likely facilitates rapid The enzyme

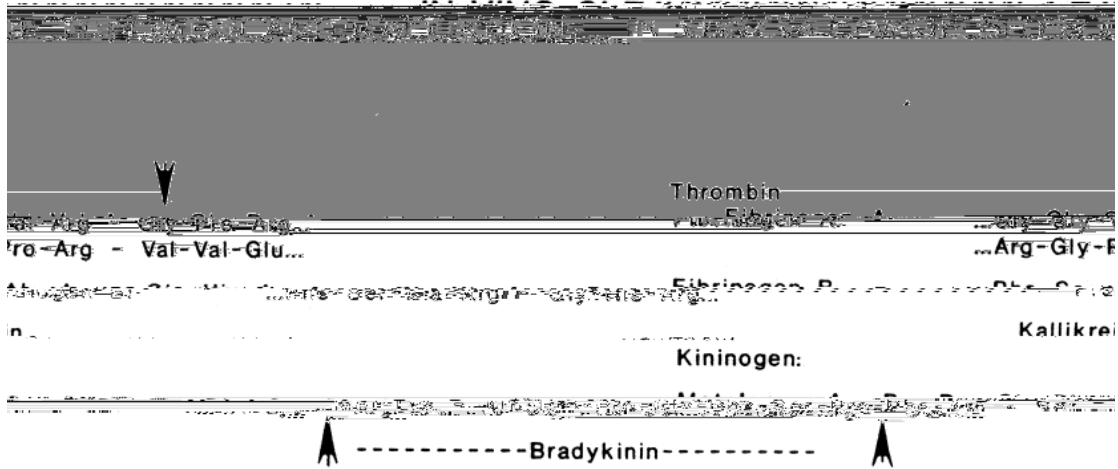


Figure 10. Several of the *in vivo* drug sites for cardiovascular modification. Paracrine and autocrine actions are based on these recognition sequences.

Proteolytic beginning toward the elucidation of these Bjarnason, J. B., and Fox, J. W. (1983).
 roles
 toxin e, a zinc protease isolated from the venom
 (Crotalus atrox). *Biochemistry* 22, 3770-3778

