

Research paper

Structural and functional characterization of complex formation between two Kunitz-type serine protease inhibitors from Russell's Viper venom

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article info

Article history:

Received 30 January 2016

Accepted 10 August 2016

Available online 12 August 2016

Keywords:

Kunitz-type serine protease inhibitor

Protease

Phospholipase A₂

Snake venom complex

Protein-protein interaction

abstract

Snake venom Kunitz-type serine protease inhibitors (KSPIs) exhibit various biological functions including anticoagulant activity. This study elucidates the occurrence and subunit stoichiometry of a putative complex formed between two KSPIs (Rusvikunin and Rusvikunin-II) purified from Russell's Viper venom.

complex was insufficiently characterized in structural terms, and little evidence was presented to support a structural association of the components of the complex. Furthermore, our studies also suggested that the Rusvikunin complex contributes to the overall toxicity of RV bites [9], thus warranting further characterization. Therefore, in this study we identify the protein components of the Rusvikunin complex by LC-MS/MS analysis. Furthermore, spectro-
uorometric, classical gel-

3. Results and discussion

Fractionation of Rusvikunin complex (see [Supplementary Fig. S1](#)) on a RP- C₁₈ HPLC column resulted in separation into two protein peaks - one large peak, named Rusvikunin-II [\[9\]](#), and a small peak, named Rusvikunin [\[8\]](#)

α_2 e macroglobulin also inhibited the brinogenolytic activity of Rusvikunin complex ([supplementary Table SII](#))

filtration column which was equilibrated with buffer containing Rusvikunin-II resulted in appearance of three major protein peaks with retention times of 11.7 min, 12.1 min and 12.3 min with corresponding molecular mass of ~21 kDa, ~14 kDa and ~7 kDa, respectively (Fig. 4A). The intensity of these equilibrium gel-filtration peaks was dependent on the injected Rusvikunin: Rusvikunin-II ratio (Fig. 4A). The highest intensity of the two major peaks eluted from the GF column with a retention time of 11.7 min

Additionally, this technique is quite efficient for studying protein-protein interactions in solution [25,26], and measurement of CD in the far UV region (178 e 260 nm) can provide valuable information on changes in the conformations of proteins when they interact to form a complex [26]. The CD spectra of Rusvikunin and Rusvikunin-II demonstrated predominance of β -sheet structure (~60%) which is in accordance to our previous observations [8,9]. Interaction of Rusvikunin with Rusvikunin-II at 1:1 and at 1:2 molar ratio did not show any significant change in the secondary structure of interacting proteins (Fig. 5). These data support the earlier observations that protein-protein interactions may result in only

3.6. Determination of interaction between Rusvikunins in solution

Acknowledgements

The authors acknowledge Prof. S.S. Ghosh, IIT, Guwahati, and the technical service provided by C-CAMP/NCBS MS Facility, Bangalore, India for helping with the CD study and LC-MS/MS analyses, respectively. SD and BK are recipients of DST-INSPIRE SRF and DBT project JRF, respectively. Financial support was received to AKM from the Department of Biotechnology, New Delhi sponsored Unit of Excellence in Biotechnology in NER of India grant (BT/412/NE/U-Excel/2013) and DBT National Bioscience Award grant (BT/HRD/NBA/34/01/2012-13/(ix)). The instrumental support received under DST-FIST grant to the Department of MBBT, TU is duly acknowledged.

Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.biochi.2016.08.005>.

References

- [1] A.K. Mukherjee, S.K. Ghosal, C.R. Maity, Some biochemical properties of Russell's viper (*Daboia russelli*) venom from Eastern India: correlation with clinical pathological manifestation in Russell's viper bite, *Toxicon* 38 (2000) 163e175.
- [2] J.R. Kumar, B.S. Basavarajappa, O. Arancio, I. Aranha, N.S. Gangadhara, H.N. Yajurvedi, T.V. Gowda, Isolation and characterization of "Reprotxin", a novel protein complex from *Daboia russelli* snake venom, *Biochimie* 90 (2008) 1545e1559.
- [3] A.K. Mukherjee, Non-covalent interaction of phospholipase A₂ (PLA₂) and kaouthiotoxin (KTX) from venom of *Naja kaouthia* exhibits marked synergism to potentiate their cytotoxicity on target cells, *J. Venom. Res.* 1 (2010) 37 e42.
- [4] R. Thakur, P. Chattopadhyay, A.K. Mukherjee, Biochemical and pharmacological characterization of a toxic fraction and its cytotoxin-like component isolated from Russell's viper (*Daboia russelli russelii*) venom, *Comp. Biochem. Physiol. Part C* 168 (2015) 5e7.
- [5] E. Delot, C. Bon, Model for the interaction of crotoxin, a phospholipase A₂ neurotoxin, with presynaptic membranes, *Biochemistry* 32 (1993) 10708e10713.
- [6] C. Bon, J.P. Changeux, T.W. Jeng, H. Fraenkel-Conrat, Postsynaptic effects of crotoxin and of its isolated subunits, *Eur. J. Biochem.* 99 (1979) 471 e481.
- [7] V. Choumet, B. Saliou, L. Fideler, Y.C. Chen, F. Gubensek, C. Bon, E. Delot, Snake-venom phospholipase A₂ neurotoxins. Potentiation of a single-chain neurotoxin by the chaperon subunit of a two-component neurotoxin, *Eur. J. Biochem.* 211 (1993) 57 e62.
- [8] A.K. Mukherjee, S.P. Mackessy, S. Dutta, Characterization of a Kunitz-type protease inhibitor peptide (Rusvikunin) purified from *Daboia russelii russelii* venom, *Int. J. Biol. Macromol.* 67 (2014) 154 e162.
- [9] A.K. Mukherjee, S.P. Mackessy, Pharmacological properties and pathophysiological significance of a Kunitz-type protease inhibitor (Rusvikunin-II) and its protein complex (Rusvikunin complex) purified from *Daboia russelii russelii* venom, *Toxicon* 89 (2014) 55 e66.
- [10] A.K. Mukherjee, B. Kalita, R. Thakur, Two acidic, anticoagulant PLA₂ isoenzymes purified from the venom of monocled cobra *Naja kaouthia* exhibit different potency to inhibit thrombin and factor Xa via phospholipids independent, non-enzymatic mechanism, *PLoS One* 9 (2014) e101334.
- [11] A.K. Mukherjee, S. Dutta, S.P. Mackessy, A new C-type lectin (RVsnaclec) purified from venom of *Daboia russelii russelii* shows anticoagulant activity via inhibition of FXa and concentration-dependent differential response to platelets in a Ca²⁺-independent manner, *Thromb. Res.* 134 (2014) 1150e1156.
- [12] A.K. Mukherjee, S.P. Mackessy, Biochemical and pharmacological properties of a new thrombin-like serine protease (Russelobin) from the venom of Russell's Viper (*Daboia russelii russelii*) and assessment of its therapeutic potential, *Biochem. Biophys. Acta Gen. Sub.* 1830 (2013) 3476e3488.
- [13] A.K. Mukherjee, The pro-coagulant brinogenolytic serine protease isoenzymes from *Daboia russelii russelii* venom coagulate the blood through factor V activation: role of glycosylation on enzymatic activity, *PLoS One* 9 (2014) e86823.
- [14] A.K. Mukherjee, A major phospholipase A₂ from *Daboia russelii russelii* venom shows potent anticoagulant action via thrombin inhibition and binding with plasma phospholipids, *Biochimie* 99 (2014) 153 e161.
- [15] R. Doley, G.F. King, A.K. Mukherjee, Differential hydrolysis of erythrocyte and mitochondrial membrane phospholipids by two phospholipase A₂ isoenzymes (NK-PLA₂-I and NK-PLA₂-II) from the venom of the Indian monocled cobra *Naja kaouthia*, *Arch. Biochem. Biophys.* 425 (2004) 1 e13.
- [16] A.P. West, A.M. Giannetti, A.M. Herr, M.J. Bennett, J.S. Nangiana, J.R. Pierce, L.P. Weiner, P.M. Snow, P.J. Bjorkman, Mutational analysis of the transferrin receptor reveals overlapping HFE and transferrin binding sites, *J. Mol. Biol.* 313 (2001) 385 e397.
- [17] S.W. Provencher, P. Stepanek, Global analysis of dynamic light scattering autocorrelation functions, *Part Part Syst. Charact.* 13 (1996) 291 e294.
- [18] A.L. Harvey, R.C. Hider, F. Khader, Effect of phospholipase A on action of cobra venom cardiotoxins on erythrocytes and skeletal muscles, *Biochim. Biophys. Acta* 728 (1983) 215 e221.
- [19] D. Saikia, R. Thakur, A.K. Mukherjee, An acidic phospholipase A₂ (RVVA-PLA₂-I) purified from *Daboia russelii* venom exerts its anticoagulant activity by enzymatic hydrolysis of plasma phospholipids and by non-enzymatic inhibition of factor Xa in a phospholipids/Ca²⁺ independent manner, *Toxicon* 57 (2011) 841 e850.
- [20] J. Liu, J. Tian, W. He, J. Xie, Z. Hu, X. Chen, Spectroscopic study of the binding of daphnetin to bovine serum albumin, *J. Pharma Biomed. Anal.* 35 (2004) 671 e677.
- [21] R. Thakur, A. Kumar, B.6m4f.73sear, D.aaar, D. Saikia,7-339.P(A)-730.6(Chattopta